

Synthesis of a New C(1 → 2)-Linked Iminodisaccharide Starting from Levoglucosenone

by Isabel Navarro and Pierre Vogel*

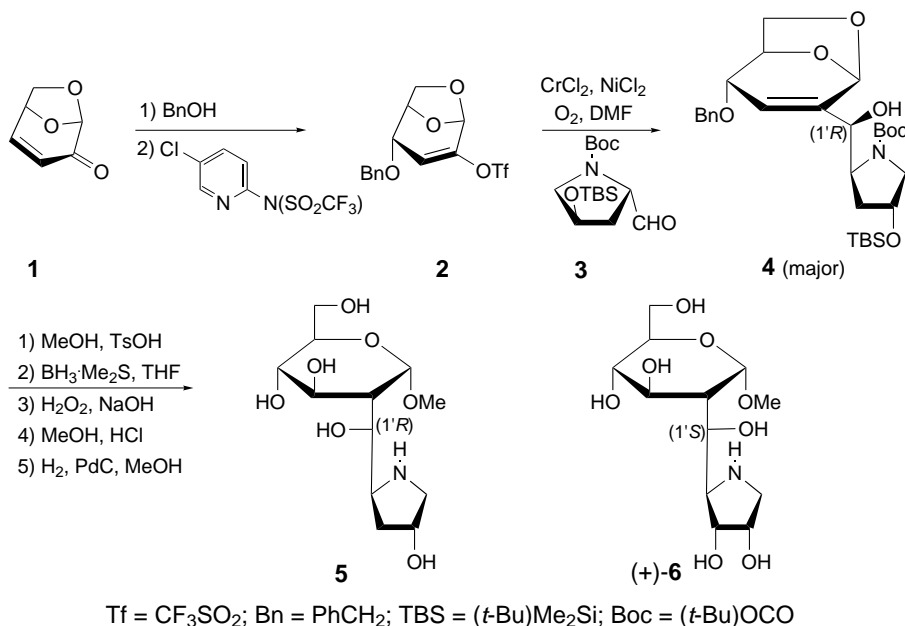
Institut de chimie organique, Ecole Polytechnique Fédérale de Lausanne, BCH, CH-1015 Lausanne-Dorigny

Methyl 2-deoxy-2-[(1S)-2,5-dideoxy-2,5-imino-L-ribitol-1-C-yl]- α -D-glucopyranoside ((+)-**6**) was obtained from the product of *Nozaki-Kishi* coupling of 2,5-[(*tert*-butoxy)carbonyl]imino-2,5-dideoxy-3,4-*O*-isopropylidene-L-ribose ((-)-**9**) and 4-*O*-benzyl-6-*O*-[(benzyloxy)methyl]-3-deoxy-2-*O*-[(trifluoromethyl)sulfonyl]- α -D-*erythro*-hex-2-enopyranoside ((+)-**12**). The alkenyl triflate (+)-**12** was derived from levoglucosenone (**1**).

Introduction. – Carbohydrate mimics are potentially useful molecular tools for biology [1] and may become leads for drug discovery [2]. In particular, C-linked disaccharides and oligosaccharides containing them offer the advantage of being resistant to acidic and enzymatic hydrolysis [3]. They are potential inhibitors of glycosidases and glycosyltransferases [4][5] and potential antibacterial, antiviral, antimetastatic, antidiabetic, antihyperglycemic, antiadhesive, and immunostimulatory agents [6][7]. A new class of selective glycosidase inhibitors has emerged, namely C-linked iminodisaccharides (aza-C-disaccharides) [8][9], which contain not only the steric and charge information of the glycosyl moiety liberated during the enzyme-catalyzed hydrolysis, but also that of the aglycon. The first example of a C-linked iminodisaccharide (1,5-dideoxy-1,5-imino-D-mannitol linked at C(6) of D-galactose through a CH₂ unit) was prepared by *Johnson et al.* [10]. Other examples of ‘linear’ C-linked iminodisaccharides were obtained by the groups of *Martin et al.* [11] and *van Boom* and co-workers [12]. We have prepared the first examples of ‘branched’ disaccharides [8][13]. Further examples were reported by *Johnson et al.* [9] and by our group [14][15]. *Brandi* and co-workers [16] have obtained the first examples of (1 → 2)-linked pseudo imino-C-disaccharides in which pyrrolidine-2,3-diol or a pyrrolidin-2-ol is linked at C(2) of D-glucose *via* a C–C bond.

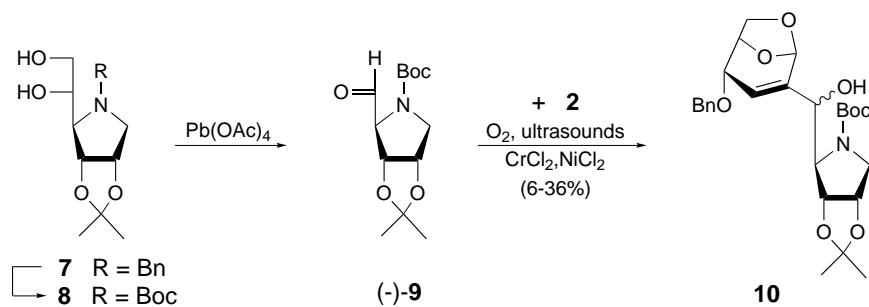
In a preliminary report [17], our group had shown that (1 → 2)-C-linked disaccharides can be obtained applying the *Nozaki-Kishi* coupling reaction to triflate **2** (derived in two steps from levoglucosenone **1** [18]) and aldehyde **3** (derived from (4*R*)-4-hydroxy-L-proline). This generated allylic alcohol **4** (48%) that was converted (*Scheme 1*) to the imino-C-disaccharide **5**. We have now applied this method to the preparation of the new C-linked iminodisaccharide (+)-**6** in which a pyrrolidine-3,4-diol moiety is attached at C(2) of methyl α -D-glucopyranoside through a hydroxymethylene linker. As we shall see, the *Nozaki-Kishi* coupling was more difficult in this case and required a monocyclic alkenyl triflate rather than the bicyclic triflate **2** for a reasonable yield of condensation. Furthermore, the diastereoselectivity of the coupling was opposite to that reported with **2** + **3** → **4** [17].

Scheme 1



Results and Discussion. – The starting aldehyde (–)-**9** was derived from the known 1,4-(benzylimino)-1,4-dideoxy-2,3-*O*-isopropylidene-D-allitol (**7**) [19]. Hydrogenolysis (H₂, Pd(OH)₂/C) in MeOH containing an excess of (*t*-BuOCO)₂O ((Boc)₂O) provided **8** (86%) [19] that was oxidized with Pb(OAc)₄/NaHCO₃ (CH₂Cl₂, –78°) to aldehyde (–)-**9** in 80% yield (Scheme 2). After a large number of unsuccessful *Nozaki-Kishi* coupling [20] attempts with triflate **2** and aldehyde (–)-**9**, we found that a 2:1 diastereoisomer mixture **10** of allylic alcohols was formed in mediocre yield (6–36%) in the presence of *ca.* 5 mol-% of O₂ and under activation with ultrasound. This suggested that the *Nozaki-Kishi* coupling is highly sensitive to steric factors: aldehyde (–)-**9** is more hindered and less flexible than aldehyde **3**.

Scheme 2



We then decided to convert the bicyclic triflate **2** into monocyclic derivative (+)-**12** (Scheme 3). Acidic methanolysis (MeOH/FB₃·Et₂O) of the anhydrohexose **2** provided (+)-**11** in 67% yield. Protection of the primary-alcohol moiety of (+)-**11** as a (benzyloxy)methyl (BOM) ether under standard conditions [21] provided (+)-**12** (68%). Its *Nozaki-Kishi* coupling with aldehyde (–)-**9** furnished as a single diastereoisomer the allylic alcohol (+)-**13**, isolated in 48% yield. In contrast to the coupling **2** + **3** → **4** (Scheme 1) for which the (1'*R*)-alcohol was obtained as major product, the reaction (–)-**9** + (+)-**12** → (+)-**13** generated a (1'*S*)-alcohol (for configuration assignment, see below). This implies that the *Re* face of aldehyde **3** is preferred for the addition of the alkenylchromium reagent derived from **2**, whereas the *Si* face of aldehyde (–)-**9** is preferred for the addition of the alkenylchromium reagent derived from (+)-**12** (see Fig. 1). The reason for this change in diastereoselectivity is unknown to us at the moment.

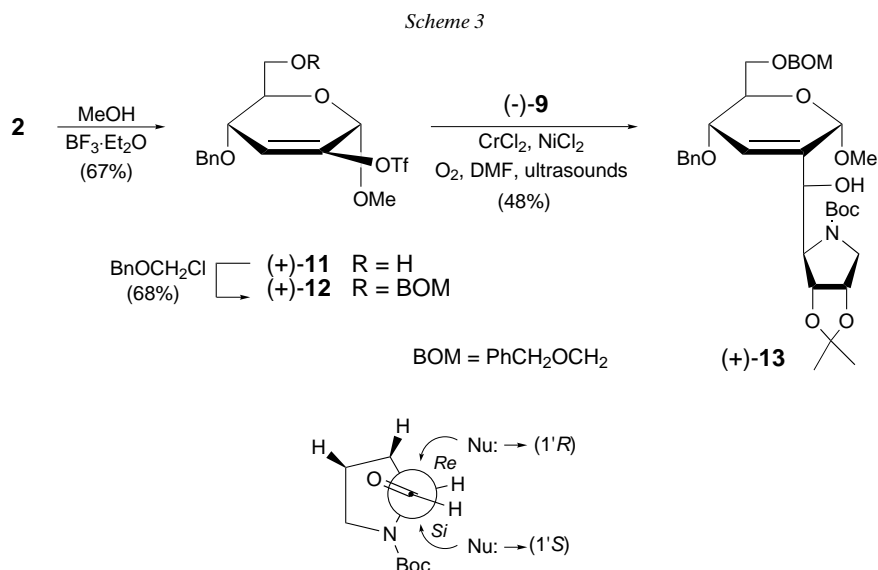
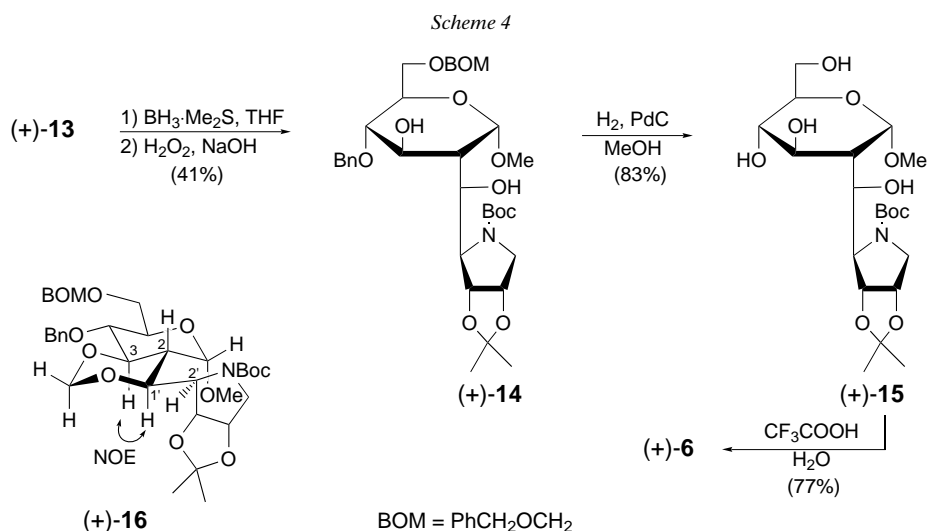


Fig. 1. Possible nucleophilic additions to aldehydes **3** and (+)-**12** (Felkin-Anh model [22])

Hydroboration of the alkene moiety of (+)-**13** with BH₃·Me₂S in THF, followed by oxidative workup (35% H₂O₂/NaOH) gave alcohol (+)-**14** in modest yield (41%) (Scheme 4). Attempts to hydroborate (+)-**13** with SmI₂/catecholborane [23], catecholborane/[RhCl(PPh₃)₃] [24] and dicyclohexylborane [25] were not met with success. As (+)-**13** was hydroborated very slowly, competitive decomposition occurred. Debenzylation of (+)-**14** (H₂, 10% Pd/C, MeOH) provided (+)-**15** (83%) that was further deprotected to give the methyl α-D-glucopyranoside (+)-**6** (77%) upon acidic treatment (CF₃COOH/H₂O 4 : 1).

The *gluco* configuration of (+)-**14**, (+)-**15**, and (+)-**6** was given by their ¹H-NMR (1D, 2D-NOESY, and COSY) data (see *Exper. Part*). The (1'*S*) configuration at the hydroxymethylene linker was determined by the ¹H-NMR and 2D-NOESY data of the dioxane derivative (+)-**16** obtained by treatment of diol (+)-**14** with CH₂Br₂ under



basic conditions (50% NaOH solution, Bu₄NBr, 60°) [26]. Typical $^3J(2,3) = 12.0$ Hz and $^3J(1',2) = 9.8$ Hz were measured. Furthermore, a strong NOE was observed for the signal pair at δ 4.36 and 4.40 assigned to H–C(1') and H–C(3), respectively.

Conformational Analysis. – The 600-MHz ¹H-NMR spectra of (+)-6 confirmed the chair conformation of the methyl α -D-glucopyranoside moiety (see *Exper. Part*). It showed a large vicinal coupling constant between protons H–C(2') and H–C(3') ($^3J = 12.8$ Hz) of the pyrrolidine ring, suggesting the envelope configuration shown in *Fig. 2*.

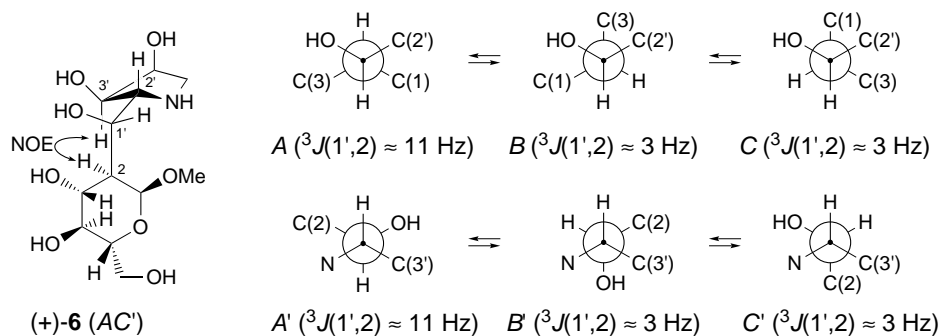


Fig. 2. Possible conformations for (+)-6

The ¹H-NMR spectra of (+)-6 measured in (D₅)pyridine at 25° showed similar vicinal coupling constants $^3J(1',2) = ^3J(1',2') = 6.8$ Hz for the proton at δ 4.60 assigned to the hydroxymethylene linker. In D₂O, the coupling constant could be measured for $^3J(1',2)$ only and was somewhat smaller (5.2 Hz) than in (D₅)pyridine. These data are consistent with equilibria of rotamers about the bond $\sigma(C(1'),C(2))$ (rotamers A, B,

and C) and the bond $\sigma(C(1'),C(2'))$ (rotamers A' , B' , and C'). Conformers A and A' (for which vicinal coupling constants ${}^3J(1',2)$ and ${}^3J(1',2')$ of ca. 11 Hz are expected [15]) must be populated at 40 to 50%. The 2D-NOESY ${}^1\text{H-NMR}$ spectra of (+)-**6** showed a intense cross-peak for proton pair $\text{H}-\text{C}(3')$ (δ 4.70) and $\text{H}-\text{C}(2)$ (δ 2.75), as expected for the conformer AC' represented in Fig. 2.

Preliminary enzymatic assays with (+)-**6** showed weak inhibitory activity toward α -mannosidases from jack bean (39% at 1 mM concentration) and from almond (26% at 1 mM concentration) [27]. Further enzymatic assays will be undertaken and will be reported in due course together with the synthesis of other C-linked iminodisaccharides.

Conclusion. – The *Nozaki-Kishi* coupling of sugar-derived alkenyl triflate with iminosugar-derived aldehydes was used to generate new C-linked iminodisaccharides. The yield and the diastereoselectivity of the coupling strongly depends on the nature of the alkenyl triflates and aldehydes. This method [17] allowed us to prepare for the first time a disaccharide mimetic in which a (2*R*,3*R*,4*S*)-3,4-dihydroxypyrrolidin-2-yl moiety (imitates α -mannosides) is attached at C(2) of methyl α -D-glucopyranoside through a hydroxymethylene linker.

This work was supported by the *Swiss National Science Foundation* (Grant No. 20,55567.98), the *Office Fédéral de l'Education et de la Science* (COST D13/0001/99), and the *Fonds Herbette* (Lausanne). We are grateful also to Mr. *Martial Rey*, *Francisco Sepúlveda*, and *Alejandro Battaner-Dubois* for technical help.

Experimental Part

General. Anh. solvents and reagents were freshly distilled under N_2 prior to use: Et_2O and THF from sodium and benzophenone; MeOH from magnesium; CH_2Cl_2 , $(\text{Me}_3\text{Si})_2\text{NH}$, iPr_2NET , pyridine, and Et_3N from CaH_2 . TLC: *Merck* silica gel 60 F_{254} plates. Flash column chromatography (FC): silica gel 60 (*Merck*, 230–400 mesh). Optical rotations: *JASCO DIP-370* digital polarimeter; at 25°; c in g/100 ml. UV Spectra: *Kontron-Uvikon 810 CW* spectrophotometer; λ_{max} in nm, ϵ in $\text{m}^{-1}\text{cm}^{-1}$. IR Spectra: *Perkin-Elmer Paragon-1000 FT-IR* spectrometer; in cm^{-1} . NMR Spectra: *Bruker DPX-400 FT* and *Bruker ARX-400 FT*, ${}^{13}\text{C}$ at 100.6 MHz; chemical shift δ in ppm, J in Hz; ${}^1\text{H}$ -assignment confirmed by 2D-COSY or/and NOESY when necessary. Mass spectra: *Nermag R-10-10C* mass spectrometer; chemical ionization (CI) mode; in m/z (rel. %).

2,5-[[*tert*-Butoxy]carbonyl]imino]-2,5-dideoxy-3,4-O-isopropylidene-L-ribose ((-)-**9**). NaHCO_3 (0.55 g, 6.54 mmol) and $\text{Pb}(\text{OAc})_4$ (2.45 g, 5.53 mmol) were added to a soln. of diol **8** [19] (1 g, 3.30 mmol) in CH_2Cl_2 (25 ml) at -78° . After 50 min at -78° , the mixture was treated with a sat. aq. NaHCO_3 soln. (50 ml) and extracted with CH_2Cl_2 (3×50 ml). The combined org. extracts were washed with brine, dried (Na_2SO_4), and evaporated. FC (light petroleum ether/AcOEt 3:7) gave 1.4 g (80%) of (-)-**9**. Colorless oil. $[\alpha]_{\text{D}}^{25} = -96$, $[\alpha]_{\text{D}}^{27} = -106$, $[\alpha]_{\text{D}}^{36} = -116$, $[\alpha]_{\text{D}}^{35} = -222$, $[\alpha]_{\text{D}}^{35} = -291$ ($c = 1.0$, CHCl_3). UV (MeCN): 198 (2340). IR (KBr): 2980, 2935, 1735, 1695, 1400, 1170, 1120, 1055, 855, 770. ${}^1\text{H-NMR}$ (400 MHz, C_6D_6): 9.32 (*s*, $\text{H}-\text{C}(1)$); 4.47 (*m*, $\text{H}-\text{C}(2)$); 4.42 (*m*, $\text{H}-\text{C}(3)$); 4.10 (*m*, $\text{H}-\text{C}(4)$); 3.89 (*d*, ${}^2J = 12.8$, $\text{H}_a-\text{C}(5)$); 3.18 (*dd*, ${}^2J = 12.8$, ${}^3J(4,5_b) = 4.8$, $\text{H}_b-\text{C}(5)$); 1.47 (*s*, Me_3C); 1.41, 1.16 (2*s*, Me_2C). ${}^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): 197.9 (*s*, CO); 154.7 (*s*, NCOO); 112.4 (*s*, Me_2C); 80.7 (*s*, Me_3C); 79.6 (*d*, $J = 145$, C(3)); 78.6 (*d*, $J = 144$, C(4)); 71.8 (*d*, $J = 144$, C(2)); 52.1 (*t*, $J = 145$, C(5)); 28.1 (*q*, $J = 143$, Me_3C); 26.7, 24.7 (2*q*, $J = 143$, Me_2C). CI-MS (NH_3): 84 (29), 142 (91), 172 (63), 186 (44), 216 (100), 233 (68), 272 (78, $[M + 1]^+$). Anal. calc. for $\text{C}_{13}\text{H}_{21}\text{NO}_5$ (271.31): C 57.55, H 7.80, N 5.16; found: C 57.62, H 7.83, N 5.10.

1,6-Anhydro-4-O-benzyl-2-[[*tert*-butoxy]carbonyl]imino]-2,5-dideoxy-3,4-O-isopropylidene-L-ribose-1-C-yl]-2,3-dideoxy- β -D-erythro-hex-2-enopyranose (**10**). A soln. of **2** [17] (0.45 g, 1.22 mmol) and (-)-**9** (0.25 g, 0.92 mmol) in anh. DMF (2 ml) was added to a mixture of CrCl_2 (0.66 g, 5.37 mmol) and NiCl_2 (3 mg, 0.023 mmol) under N_2 . After injection of O_2 (5 ml, 0.22 mmol), the flask was sealed and sonicated at 20° for 1 h. The mixture was diluted with light petroleum ether/AcOEt 1:1. After the addition of 1M sodium serinate, the mixture was stirred vigorously at 20° for 2 h and extracted with CH_2Cl_2 (3×50 ml). The combined

org. phases were washed with brine, dried (Na_2SO_4), and evaporated. FC (light petroleum ether/AcOEt 3:2) gave 0.16 g (36%) of **10**, 2:1 diastereoisomer mixture. Colorless oil. UV (MeCN): 248 (1530), 233 (1200). IR (film): 3340, 2985, 1695, 1415, 1165, 1125, 1050, 985, 905. $^1\text{H-NMR}$ (400 MHz, CDCl_3 ; major isomer): 7.37 (*m*, Ph); 5.82 (*m*, H–C(3)); 5.64 (*s*, H–C(1)); 4.69 (*m*, PhCH_2 , H–C(5), H–C(1')); 4.20–3.64 (*m*, H–C(3'), H–C(4'), H–C(2'), $\text{H}_{\text{exo}}-\text{C}(6)$, H–C(4), $\text{H}_a-\text{C}(5')$); 3.49 (*dd*, $^2J=12.4$, $^3J(4',5'b)=5.2$, $\text{H}_b-\text{C}(5')$); 3.38 (*m*, $\text{H}_{\text{endo}}-\text{C}(6)$); 1.49 (*s*, Me_3C); 1.30 (*s*, Me_2C). $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3 ; major isomer): 154.9 (*s*, NCOO); 143.7 (*s*, C(2)); 138.1 (*s*, arom. C); 128.3, 127.5 (*3d*, $^1J(\text{C,H})=140$, arom. C); 118.9 (*d*, $J=68$, C(3)); 111.3 (*s*, Me_2C); 95.8 (*d*, $J=167$, C(1)); 80.1 (*s*, Me_3C); 79.4 (*d*, $J=145$, C(3')); 78.9 (*d*, $J=144$, C(4')); 74.4 (*d*, $J=144$, C(2')); 72.4 (*d*, $J=145$, C(1')); 70.2 (*t*, $J=144$, C(6)); 67.5 (*d*, $J=145$, C(4)); 67.3 (*d*, $J=145$, C(5)); 53.8 (*d*, $J=144$, C(5')); 28.3 (*q*, $J=144$, Me_3C); 26.9, 24.8 (*2q*, $J=144$, Me_2C). CI-MS (NH_3): 91 (39), 142 (89), 172 (32), 233 (100), 272 (55), 326 (17), 490 (78, $[M+1]^+$). Anal. calc. for $\text{C}_{26}\text{H}_{35}\text{NO}_8$ (489.57): C 63.79, H 7.21, N 2.86; found: C 63.90, H 7.15, N 2.91.

Methyl 4-O-Benzyl-3-deoxy-2-O-[(trifluoromethyl)sulfonyl]- α -D-erythro-hex-2-enopyranoside ((+)-11). $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (0.7 ml, 5.4 mmol) was added to a soln. of **2** [17] (0.5 g, 1.4 mmol) in MeOH (4 ml) at 0° . The mixture was left at 20° overnight. Evaporation and FC (light petroleum ether/AcOEt 1:1) gave 0.36 g (67%) of (+)-**11**. White solid. M.p. $85-86^\circ$. $[\alpha]_{\text{D}}^{25} = +102$, $[\alpha]_{\text{D}}^{27} = +130$, $[\alpha]_{\text{D}}^{25} = +142$, $[\alpha]_{\text{D}}^{25} = +152$, $[\alpha]_{\text{D}}^{25} = +163$ ($c=0.1$, CHCl_3). UV (MeCN): 218 (19480), 206 (5380). IR (KBr): 3855, 3445, 3005, 1715, 1635, 1495, 1390, 1140, 910, 700. $^1\text{H-NMR}$ (400 MHz, CDCl_3): 7.39 (*m*, arom. C); 6.07 (*d*, $^3J(4,3)=2.0$, H–C(3)); 4.91 (*s*, H–C(1)); 4.62 (*AB*, $^2J=11.6$, PhCH_2); 4.35 (*dd*, $^3J(5,4)=9.6$, $^3J(3,4)=2.0$, H–C(4)); 3.92 (*ddd*, $^3J(4,5)=9.6$, $^3J(6b,5)=3.6$, $^3J(6a,5)=1.6$, H–C(5)); 3.38 (*dd*, $^2J=11.6$, $^3J(5,6a)=1.6$, $\text{H}_a-\text{C}(6)$); 3.37 (*dd*, $^2J=11.6$, $^3J(5,6b)=3.6$, $\text{H}_b-\text{C}(6)$); 3.48 (*s*, MeO). $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): 145.0 (*s*, CF_3); 137.1 (*s*, C(2)); 137.2 (*s*, arom. C); 128.6, 128.3, 127.9 (*3d*, $J=161$, arom. C); 119.5 (*d*, $J=168$, C(3)); 94.6 (*d*, $J=167$, C(1)); 71.5 (*d*, $J=145$, C(5)); 70.1 (*t*, $J=143$, PhCH_2); 69.9 (*d*, $J=144$, C(4)); 61.3 (*t*, $J=144$, C(6)); 56.5 (*q*, $J=144$, MeO). $^{19}\text{F-NMR}$ (376 MHz, CDCl_3): -77.6 (*s*, CF_3). CI-MS (NH_3): 91 (64), 108 (25), 338 (9), 384 (22), 415 (76, $[M+\text{NH}_3]^+$). Anal. calc. for $\text{C}_{15}\text{H}_{17}\text{F}_3\text{O}_8$ (398.01): C 45.23, H 4.30, S 8.05; found: C 45.33, H 4.40, S 7.98.

Methyl 4-O-Benzyl-6-O-[(benzyloxy)methyl]-3-deoxy-2-O-[(trifluoromethyl)sulfonyl]- α -D-erythro-hex-2-enopyranoside ((+)-12). At 0° , (+)-**11** (0.36 g, 0.91 mmol) and $^i\text{Pr}_2\text{NH}$ (0.2 ml, 1.2 mmol) were mixed with benzyl chloromethyl ether (0.2 ml, 1.2 mmol). The mixture was allowed to stand at 20° overnight. Then 1*n* aq. HCl (1 ml) was added, the org. phase separated, dried, (Na_2SO_4) and evaporated. FC (light petroleum ether/AcOEt 4:1) gave 0.32 g (68%) of (+)-**12**. Colorless oil. $[\alpha]_{\text{D}}^{25} = +36$, $[\alpha]_{\text{D}}^{27} = +48$, $[\alpha]_{\text{D}}^{25} = +52$, $[\alpha]_{\text{D}}^{25} = +123$, $[\alpha]_{\text{D}}^{25} = +175$ ($c=0.1$, CHCl_3). UV (MeCN): 218 (18000), 209 (4300). IR (film): 3440, 3005, 1715, 1635, 1485, 1490, 1380, 1140, 900, 700. $^1\text{H-NMR}$ (400 MHz, CDCl_3): 7.35 (*m*, arom. H); 6.09 (*d*, $^3J(4,3)=2.4$, H–C(3)); 4.95 (*s*, H–C(1)); 4.79 (*AB*, $^2J=11.6$, PhCH_2); 4.64 (*m*, CH_2); 4.41 (*dd*, $^3J(4,5)=9.6$, $^3J(3,4)=2.0$, H–C(4)); 4.04 (*ddd*, $^3J(4,5)=9.6$, $^3J(6b,5)=3.6$, $^3J(6a,5)=1.6$, H–C(5)); 3.81 (*m*, H–C(6)); 3.48 (*s*, MeO). $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): 145.0 (*s*, CF_3); 138.3 (*s*, C(2)); 137.1 (*s*, arom. C); 129.4, 129.5, 129.0, 128.7, 127.8, 127.4 (*5d*, $J=161$, arom. C); 119.5 (*d*, $J=168$, C(3)); 94.6 (*d*, $J=167$, C(1)); 94.5 (*t*, $J=167$, OCH_2O); 70.1 (*t*, $J=143$, PhCH_2); 69.9 (*d*, $J=144$, C(4)); 68.3 (*d*, $J=145$, C(5)); 65.4 (*t*, $J=143$, CH_2); 61.3 (*t*, $J=144$, C(6)); 56.5 (*q*, $J=144$, MeO). $^{19}\text{F-NMR}$ (376 MHz, CDCl_3): -77.6 (*s*, CF_3). CI-MS (NH_3): 91 (62), 108 (25), 217 (14), 535 (100, $[M+\text{NH}_3]^+$). Anal. calc. for $\text{C}_{25}\text{H}_{25}\text{F}_3\text{O}_8\text{S}$ (518.03): C 53.25, H 4.82, S 6.17; found: C 53.26, H 4.89, S 6.13.

*Methyl 4-O-Benzyl-6-O-[(benzyloxy)methyl]-2-[(1*S*)-2,5-[(tert-butoxy)carbonyl]imino]-2,5-dideoxy-3,4-O-isopropylidene-L-ribitol-1-C-yl]-3-deoxy- α -D-erythro-hex-2-enopyranoside ((+)-13)*. As described for **10**, with (+)-**12** (0.27 g, 0.52 mmol), (–)-**9** (0.1 g, 0.37 mmol), DMF (2 ml), CrCl_2 (0.33 g, 2.68 mmol), NiCl_2 (2.6 mg, 0.012 mmol), and O_2 (3 ml, 0.14 mmol). FC (light petroleum ether/AcOEt 1:1) gave 0.11 g (48%) of (+)-**13**. Colorless oil. $[\alpha]_{\text{D}}^{25} = +229$, $[\alpha]_{\text{D}}^{27} = +234$, $[\alpha]_{\text{D}}^{25} = +248$, $[\alpha]_{\text{D}}^{25} = +312$, $[\alpha]_{\text{D}}^{25} = +456$ ($c=0.8$, CHCl_3). UV (MeCN): 218 (16600), 260 (2212). IR (film): 3370, 2985, 1710, 1655, 1435, 1550, 1105, 1015, 970, 855, 775, 705. $^1\text{H-NMR}$ (400 MHz, CDCl_3): 7.34–7.28 (*m*, arom. H); 6.01 (*m*, H–C(3)); 5.12 (*s*, H–C(1)); 4.81, 4.78 (*AB*, $^2J=16.4$, PhCH_2); 4.69 (*m*, H–C(4), H–C(5), $\text{H}_a-\text{C}(6)$); 4.63 (*s*, CH_2O); 4.42 (*dd*, $^2J=11.2$, $^3J(5,6b)=3.6$, $\text{H}_b-\text{C}(6)$); 4.04 (*m*, H–C(1')); 4.14 (*m*, H–C(2')); 3.48 (*s*, MeO); 3.95 (*d*, $^2J=12.0$, $\text{H}_a-\text{C}(5')$); 3.86 (*m*, H–C(3')); 3.81 (*dd*, $^3J(3',4')=^3J(5'b,4')=4.8$, H–C(4')); 3.42 (*dd*, $^2J=12.0$, $^3J(4',5'b)=4.8$, $\text{H}_b-\text{C}(5')$); 1.48 (*s*, Me_3C); 1.46, 1.33 (*2s*, Me_2C). $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): 154.1 (*s*, NCOO); 138.3 (*s*, C(2)); 136.5 (*s*, arom. C); 129.4, 129.5, 129.0, 128.7, 127.8, 127.4 (*6d*, $J=161$, arom. C); 124.1 (*d*, $J=168$, C(3)); 111.5 (*s*, Me_2C); 95.8 (*d*, $J=167$, C(1)); 94.5 (*t*, $J=144$, OCH_2O); 82.5 (*s*, Me_3C); 80.1 (*d*, $J=145$, C(3')); 79.0 (*d*, $J=144$, C(4')); 72.4 (*d*, $J=144$, C(2')); 71.4 (*d*, $J=145$, C(1')); 70.8 (*d*, $J=143$, CH_2O); 69.1 (*t*, $J=144$, C(6)); 68.9 (*d*, $J=144$, C(4)); 67.2 (*d*, $J=145$, C(5)); 66.3 (*d*, $J=143$, CH_2O); 55.7 (*q*, $J=144$, MeO); 53.1 (*d*, $J=144$, C(5')); 28.9, 28.5, 28.1 (*3q*, $J=144$, Me_3C); 27.3, 25.0 (*2q*, $J=144$, Me_2C). CI-MS (NH_3): 91 (40), 142 (31), 172

(31), 186 (16), 233 (100), 273 (12), 611 (63). Anal. calc. for $C_{35}H_{47}NO_{10}$ (641.04): C 65.49, H 7.33, N 2.18; found: C 65.42, H 7.45, N 2.27.

Methyl 4-O-Benzyl-6-O-[(benzyloxy)methyl]-2-[(1S)-2,5-[[tert-butoxy]carbonyl]imino]-2,5-dideoxy-3,4-O-isopropylidene-L-ribose-1-C-yl]-2-deoxy- α -D-glucopyranoside ((+)-14). A 10M $BH_3 \cdot SMe_2$ soln. (0.15 ml, 1.5 mmol) was added dropwise to a stirred soln. of (+)-13 (0.1 g, 0.15 mmol) in THF (3 ml) under N_2 cooled to 0° . The mixture was heated under reflux for 1 h. After cooling to 0° , 3M NaOH (4 ml), then 35% H_2O_2 soln. (4 ml) were added. The mixture was stirred vigorously at 20° for 1 h. Then, 1M potassium tartrate (6 ml) was added, and the mixture was extracted with CH_2Cl_2 (4×10 ml). The combined org. phases were dried (Na_2SO_4) and evaporated. FC (AcOEt/light petroleum ether 3:2) gave 41 mg (41%) of (+)-14. Colorless oil. $[\alpha]_{589}^{25} = +2$, $[\alpha]_{577}^{25} = +4$, $[\alpha]_{546}^{25} = +7$, $[\alpha]_{435}^{25} = +13$, $[\alpha]_{255}^{25} = +14$ ($c = 0.8$, $CHCl_3$). UV (MeCN): 206 (10500). IR (film): 3375, 2985, 1675, 1405, 1550, 1210, 1165, 870, 755. 1H -NMR (400 MHz, $CDCl_3$): 7.34–7.28 (*m*, arom. H); 5.26 (*d*, $^3J(2,1) = 3.5$, H–C(1)); 4.90 (*dd*, $^3J(5,4) = 5.9$, $^3J(3,4) = 3.2$, H–C(4)); 4.83, 4.79 (*AB*, $^2J = 16.4$, $PhCH_2$); 4.67 (*m*, H–C(5), H_a –C(6)); 4.61 (*s*, CH_2O); 4.57 (*m*, H_b –C(6)); 4.01 (*m*, H–C(1')); 3.86 (*m*, H_a –C(5')); 3.74 (*m*, H–C(3), H–C(3')); 3.66 (*dd*, $^3J(3',4') = ^3J(5'b,4') = 5.4$, H–C(4')); 3.47 (*dd*, $^3J(3',2') = 9.1$, $^3J(1',2') = 4.0$, H–C(2')); 3.38 (*s*, MeO); 3.22 (*dd*, $^2J = 12.0$, $^3J(4',5'b) = 4.8$, H_b –C(5')); 1.95 (*ddd*, $^3J(3,2) = 11.4$, $^3J(1',2) = 7.5$, $^3J(1,2) = 3.5$, H–C(2)); 1.45 (*s*, Me_2C); 1.32, 1.29 (2*s*, Me_2C). ^{13}C -NMR (100.6 MHz, $CDCl_3$): 155.4 (*s*, NCOO); 137.7 (*s*, arom. C); 129.4, 129.5, 129.0, 128.7, 127.8, 127.4 (6*d*, $J = 161$, arom. C); 111.3 (*s*, Me_2C); 97.7 (*d*, $J = 167$, C(1)); 94.9 (*t*, $J = 144$, OCH_2O); 80.2 (*s*, Me_3C); 80.1 (*d*, $J = 145$, C(3')); 78.9 (*d*, $J = 144$, C(4')); 72.4 (*d*, $J = 144$, C(2')); 70.8 (*d*, $J = 145$, C(1')); 70.1 (*d*, $J = 143$, CH_2O); 69.3 (*t*, $J = 144$, C(6)); 67.5 (*d*, $J = 144$, C(4)); 66.7 (*d*, $J = 145$, C(5)); 65.2 (*t*, $J = 143$, CH_2O); 63.5 (*d*, $J = 145$, C(3)); 54.2 (*d*, $J = 144$, C(5')); 52.6 (*q*, $J = 144$, MeO); 40.2 (*d*, $J = 145$, C(2)); 28.3 (*q*, $J = 144$, Me_3C); 27.3, 25.0 (2*q*, $J = 144$, Me_2C). CI-MS (NH_3): 91 (13), 142 (50), 174 (28), 186 (16), 202 (13), 218 (99), 219 (52), 235 (28), 275 (57), 661 (16). Anal. calc. for $C_{35}H_{49}NO_{11}$ (659.13): C 63.73, H 7.44, N 2.13; found: C 63.68, H 7.56, N 2.18.

Methyl 2-[(1S)-2,5-[[tert-Butoxy]carbonyl]imino]-2,5-dideoxy-3,4-O-isopropylidene-L-ribose-1-C-yl]-2-deoxy- α -D-glucopyranoside ((+)-15). A degassed vac. line mixture of (+)-14 (35 mg, 0.053 mmol), 10% Pd/C (12 mg, 0.01 mmol), and MeOH (3 ml) was stirred under H_2 at 20° overnight. The catalyst was filtered off and the solvent evaporated: 20 mg (83%) of (+)-15. White solid. M.p. 135 – 136° . $[\alpha]_{589}^{25} = +123$, $[\alpha]_{577}^{25} = +133$, $[\alpha]_{546}^{25} = +135$, $[\alpha]_{435}^{25} = +190$, $[\alpha]_{255}^{25} = +417$ ($c = 0.6$, H_2O). UV (MeCN): 199 (7680). IR (KBr): 3855, 3365, 2975, 2365, 1670, 1575, 1365, 1210, 870, 775. 1H -NMR (400 MHz, D_2O): 4.98 (*d*, $^3J(2,1) = 4.9$, H–C(1)); 4.81 (*m*, H–C(4), H–C(5), H_a –C(6), H_b –C(6)); 3.96 (*dd*, $^2J = 10.0$, $^3J(4',5'b) = 4.8$, H–C(5')); 3.37 (*m*, H–C(3')); 3.71 (*m*, H–C(4')); 3.66 (*m*, H–C(1'), H–C(2')); 3.55 (*dd*, $^2J = 10.0$, $^3J(4',5'a) = 4.4$, H_a –C(5')); 3.38 (*dd*, $^3J(2,3) = 12.7$, $^3J(4,3) = 4.8$, H–C(3)); 3.29 (*s*, MeO); 1.82 (*ddd*, $^3J(3,2) = 12.7$, $^3J(1',2) = 7.5$, $^3J(1,2) = 4.9$, H–C(2)); 1.37 (*s*, Me_3C); 1.35, 1.25 (2*s*, Me_2C). ^{13}C -NMR (100.6 MHz, D_2O): 156.4 (*s*, NCOO); 111.9 (*s*, Me_2C); 98.7 (*d*, $J = 167$, C(1)); 82.1 (*d*, $J = 145$, C(3')); 81.1 (*s*, Me_3C); 78.7 (*d*, $J = 144$, C(4')); 71.4 (*d*, $J = 144$, C(2')); 71.1 (*d*, $J = 145$, C(1')); 69.4 (*d*, $J = 144$, C(4)); 66.9 (*d*, $J = 145$, C(5)); 69.3 (*t*, $J = 144$, C(6)); 60.7 (*d*, $J = 145$, C(3)); 54.6 (*d*, $J = 144$, C(5')); 51.3 (*q*, $J = 144$, MeO); 43.2 (*d*, $J = 145$, C(2)); 27.6 (*q*, $J = 144$, Me_3C); 25.6 (*q*, $J = 144$, Me_2C). CI-MS (NH_3): 85 (25), 142 (55), 186 (30), 242 (19), 284 (7), 344 (3), 450 (3, $[M+1]^+$). Anal. calc. for $C_{20}H_{35}NO_{10}$ (449.23): C 53.43, H 7.85, N 3.12; found: C 53.35, H 7.90, N 3.83.

Methyl 2-Deoxy-2-[(1S)-2,5-dideoxy-2,5-imino-L-ribose-1-C-yl]- α -D-glucopyranoside ((+)-6). A mixture of (+)-15 (37 mg, 0.082 mmol) and 80% aq. CF_3COOH soln. (4 ml) was stirred at 20° for 2 h. The soln. was poured onto a column (1 \times 5 cm) of Dowex 50WX8 (100–200 mesh) and the column eluted sequentially with MeOH (20 ml), H_2O (5 ml), and 25% aq. NH_3 soln. (40 ml). The product fractions afforded 20 mg (77%) of (+)-6. White solid. M.p. 160° (dec.). $[\alpha]_{589}^{25} = +53$, $[\alpha]_{577}^{25} = +54$, $[\alpha]_{546}^{25} = +63$, $[\alpha]_{435}^{25} = +105$, $[\alpha]_{255}^{25} = +161$ ($c = 0.4$, H_2O). UV (MeCN): 208 (4700). IR (KBr): 3885, 2535, 1495, 1440, 1210, 1135, 1025, 835, 800, 725. 1H -NMR (600 MHz, (D_5) pyridine): 5.55 (*d*, $^3J(2,1) = 4.0$, H–C(1)); 4.70 (*dd*, $^3J(2',3') = 12.8$, $^3J(4',3') = 5.6$, H–C(3')); 4.60 (*t*, $^3J(2',1') = ^3J(2',1') = 6.8$, H–C(1')); 4.54 (*dd*, $^3J(2,3) = 10.8$, $^3J(4,3) = 7.6$, H–C(3)); 4.45 (*m*, H–C(4'), H_a –C(5'), H_a –C(6)); 4.31 (*dd*, $^2J = 12.4$, $^3J(5,6b) = 5.6$, H_b –C(6)); 4.21 (*m*, H–C(5)); 4.13 (*dd*, $^3J(5,4) = 9.6$, $^3J(3,4) = 7.6$, H–C(4)); 4.08 (*dd*, $^3J(3',2') = 12.8$, $^3J(1',2') = 6.8$, H–C(2')); 3.31 (*m*, H_b –C(5')); 3.35 (*s*, MeO); 2.75 (*ddd*, $^3J(3,2) = 10.8$, $^3J(1',2) = 6.8$, $^3J(1,2) = 4.0$, H–C(2)). ^{13}C -NMR (100.6 MHz, (D_5) pyridine): 100.4 (*d*, $J = 167$, C(1)); 74.5 (*d*, $J = 145$, C(3')); 73.9 (*d*, $J = 144$, C(4')); 73.2 (*d*, $J = 144$, C(2')); 72.7 (*d*, $J = 145$, C(1')); 71.3 (*d*, $J = 144$, C(4)); 70.8 (*d*, $J = 145$, C(5)); 66.5 (*t*, $J = 144$, C(6)); 62.8 (*d*, $J = 145$, C(3)); 54.3 (*d*, $J = 144$, C(5')); 52.2 (*q*, $J = 144$, MeO); 48.9 (*d*, $J = 145$, C(2)). CI-MS (NH_3): 102 (65), 134 (83), 180 (10), 229 (13), 256 (36), 278 (29), 310 (66, $[M+1]^+$). Anal. calc. for $C_{12}H_{23}NO_8$ (309.32): C 46.60, H 7.49, N 4.53; found: C 46.55, H 7.36, N 4.45.

Methyl 4-O-Benzyl-6-O-[(benzyloxy)methyl]-2-[(1'S)-2,5-[[tert-butoxy]carbonyl]imino]-2',5'-dideoxy-3',4'-O-isopropylidene-L-ribose-1'-C-yl]-2-deoxy-1',3'-O-methylene- α -D-glucopyranoside (= (6S)-6-(1R)-1,4-

[[tert-Butoxy]carbonyl]imino]-1,4-dideoxy-2,3-O-isopropylidene-L-erythritol-1-C-yl]-5,6-dihydro(methyl 4-O-benzyl-6-O-[(benzyloxy)methyl]-2,3-dideoxy- α -D-glucopyranosido)[2,3-d]-4H-1,3-dioxin; (+)-**16**). A mixture of (+)-**14** (32 mg, 0.048 mmol), 50% aq. NaOH soln. (0.7 ml, 0.87 mmol), Bu₄NBr (2 mg, 0.006 mmol), and CH₂Br₂ (0.19 ml, 1.1 mmol) was vigorously stirred at 55° for 2 h. Then, the mixture was extracted with CH₂Cl₂ (4 × 10 ml) and the combined org. phase dried (Na₂SO₄) and evaporated. FC (AcOEt/light petroleum ether 1:1) gave 16 mg (46%) of (+)-**16**. Colorless oil. [α]_D²⁵ = +12, [α]_D³⁵ = +13, [α]_D³⁴⁶ = +14, [α]_D³³⁵ = +23, [α]_D³⁰⁵ = +35 (*c* = 0.4, CHCl₃). UV (MeCN): 194 (2530). IR (film): 2995, 2935, 1675, 1380, 1210, 1165, 870, 505. ¹H-NMR (400 MHz, CDCl₃): 7.30–7.27 (*m*, arom. H); 5.07 (*m*, H–C(4)); 4.94 (*d*, ³*J*(2,1) = 2.5, H–C(1)); 4.92 (*m*, CH₂O); 4.83, 4.77 (*AB*, ²*J* = 16.4, PhCH₂); 4.66 (*m*, H–C(5), H_a–C(6), H_b–C(6)); 4.59 (*s*, OCH₂O); 4.40 (*dd*, ³*J*(2,3) = 12.0, ³*J*(4,3) = 3.7, H–C(3)); 4.36 (*dd*, ³*J*(2,1') = 9.8, ³*J*(2',1') = 3.4, H–C(1')); 3.85 (*dd*, ³*J*(3',2') = 9.3, ³*J*(1',2') = 3.4, H–C(2')); 3.65 (*m*, H_a–C(5')); 3.63 (*m*, H–C(4'), H–C(3'), H_b–C(5')); 3.35 (*s*, MeO); 2.57 (*ddd*, ³*J*(3,2) = 12.0, ³*J*(1',2) = 9.8, ³*J*(1,2) = 2.5, H–C(2)); 1.42 (*s*, Me₃C); 1.25 (*s*, Me₂C). ¹³C-NMR (100.6 MHz, CDCl₃): 155.4 (*s*, NCOO); 137.7 (*s*, arom. C); 129.4, 129.5, 129.0, 128.7, 127.8, 127.4 (6*d*, *J* = 161, arom. C); 110.6 (*s*, Me₂C); 97.7 (*d*, *J* = 167, C(1)); 94.9 (*t*, *J* = 144, OCH₂O); 89.3 (*t*, *J* = 144, OCH₂O); 81.8 (*s*, Me₃C); 79.8 (*d*, *J* = 145, C(3')); 79.7 (*d*, *J* = 144, C(4')); 74.6 (*d*, *J* = 144, C(2)); 74.5 (*d*, *J* = 145, C(1')); 72.2 (*d*, *J* = 145, C(3)); 70.3 (*d*, *J* = 143, CH₂O); 69.3 (*t*, *J* = 144, C(6)); 66.6 (*d*, *J* = 144, C(4)); 65.8 (*d*, *J* = 145, C(5)); 55.1 (*t*, *J* = 143, CH₂O); 53.9 (*t*, *J* = 144, C(5')); 43.5 (*q*, *J* = 144, MeO); 34.1 (*d*, *J* = 145, C(2)); 28.5 (*q*, *J* = 144, Me₃C); 26.8, 24.5 (2*q*, *J* = 144, Me₂C). ES-MS (NH₃): 434 (10), 459 (25), 540 (100), 559 (60), 640 (10), 672 (12), 720 (5, *M*⁺). Anal. calc. for C₃₉H₆₁NO₁₁ (719.92): C 65.07, H 8.54, N 1.95; found: C 64.97, H 8.15, N 1.94.

REFERENCES

- [1] See, e.g., A. Varki, *Glycobiology* **1993**, *3*, 97; T. Feizi, *Curr. Opin. Struct. Biol.* **1993**, *3*, 701; A. Varki, *Proc. Natl. Acad. Sci. U.S.A.* **1994**, *91*, 7390; K. W. Moremen, R. B. Trimble, A. Herscovics, *Glycobiology* **1994**, *4*, 113; D. J. O'Leary, Y. Kishi, *Tetrahedron Lett.* **1994**, *35*, 5591; B. Aguilera, J. Jiménez-Barbero, A. Fernandez-Mayoralas, *Carbohydr. Res.* **1998**, *308*, 19; A. Wei, K. M. Boy, Y. Kishi, *J. Am. Chem. Soc.* **1995**, *117*, 9432; J.-F. Espinosa, F. J. Cañada, J. L. Asensio, M. Martín-Pastor, H. Dietrich, M. Martín-Lomas, R. R. Schmidt, J. Jiménez-Barbero, *J. Am. Chem. Soc.* **1996**, *118*, 10862; J. F. Espinosa, E. Montero, A. Vian, J. L. García, H. Dietrich, A. Imberty, F. J. Cañada, R. R. Schmidt, J. Jiménez-Barbero, M. Martín-Lomas, *J. Am. Chem. Soc.* **1998**, *120*, 1309; A. D. Elbein, R. J. Molineaux, in 'Iminosugars as Glycosidase Inhibitors, Norijimycin and Beyond', Ed. A. E. Stütz, Wiley-VCH, Weinheim, 1998, p. 216; J. F. Espinosa, M. Bruix, O. Jarretton, T. Skrydstrup, J.-M. Beau, J. Jiménez-Barbero, *Chem.–Eur. J.* **1999**, *5*, 442.
- [2] See, e.g., M. S. Mulligan, J. C. Paulson, S. DeFrees, Z. L. Zheng, J. B. Lowe, P. A. Ward, *Nature (London)* **1993**, *364*, 149; G. D. MacLean, M. Reddish, R. Kogarty, T. Wong, S. Gandhi, M. Smolenski, J. Samuel, J. M. Nabholt, B. M. Longenecker, *Cancer Immunol. Immunother.* **1993**, *36*, 215; I. Vlahov, R. Vlahova, R. J. Linhardt, *J. Am. Chem. Soc.* **1997**, *119*, 1480; F. M. Platt, G. Reinkensmneier, R. A. Dwek, T. D. Butters, *J. Biol. Chem.* **1997**, *272*, 19365; A. Mehta, X. Lu, T. M. Block, B. S. Blumberg, R. A. Dwek, *Proc. Natl. Acad. Sci. U.S.A.* **1997**, *94*, 1822; P. Sinay, *Pure Appl. Chem.* **1997**, *69*, 459; P. Sinay, *Pure Appl. Chem.* **1998**, *70*, 407; M. Bols, *Acc. Chem. Res.* **1998**, *31*, 1; P. Sears, C.-H. Wong, *Angew. Chem., Int. Ed.* **1999**, *38*, 2301, and ref. cit. therein.
- [3] D. E. Levy, C. Tang, in 'The Chemistry of C-Glycosides', Tetrahedron Organic Chemistry Series, Eds. J. E. Baldwin and P. D. Magnus, Pergamon-Elsevier Science, Oxford, 1995; M. H. D. Postema, in 'C-Glycoside Synthesis', CRC, Boca Raton, FL, 1995; P. Vogel, R. Ferritto, K. Kraehenbuehl, A. Baudat, in 'Carbohydrate Mimics. Concept and Methods', Ed. Y. Chapleur, Wiley-VCH, Weinheim, 1998, p. 19, and ref. cit. therein; Y. Du, R. J. Linhardt, I. R. Vlahov, *Tetrahedron* **1998**, *54*, 9913.
- [4] C. Pasquarello, R. Demange, P. Vogel, *Bioorg. Med. Chem. Lett.* **1999**, *9*, 793.
- [5] C. Pasquarello, S. Picasso, R. Demange, M. Malissard, E. G. Berger, P. Vogel, *J. Org. Chem.* **2000**, *65*, 4251.
- [6] See, e.g., P. D. Rye, N. V. Bovin, E. V. Vlasova, R. A. Walker, *Glycobiology* **1995**, *5*, 385; D. K. C. Cooper, R. Oriol, 'Glycobiology in Xenotransplantation Research', in 'Glycosciences', Ed. S. Gabius, Chapman & Hall, Weinheim, 1997, p. 531; E. C. Hallberg, V. Strokan, T. D. H. Cairns, M. E. Breimer, B. E. Samuelsson, *Xenotransplantation* **1998**, *5*, 246; S. D. Kuduk, J. B. Schwarz, X.-T. Chen, P. W. Glunz, D. Sames, G. Ragupathi, P. O. Livingston, S. Danishefsky, *J. Chem. Soc., Chem. Commun.* **1998**, *120*, 12474.
- [7] See, e.g., B. Ganem, *Acc. Chem. Res.* **1996**, *29*, 340, and ref. cit. therein; L. A. G. M. van den Broek, D. J. Vermaas, B. M. Heskamp, C. A. A. van Boeckel, M. C. A. A. Tan, J. G. M. Bolscher, H. L. Ploegh, F. J. van

- Kemenade, R. E. Y. de Goede, F. Miedema, *Recl. Trav. Chim. Pays-Bas* **1993**, *112*, 82; G. C. Look, C. H. Fotsch, C.-H. Wong, *Acc. Chem. Res.* **1993**, *26*, 182; B. Winchester, G. W. J. Fleet, *Glycobiology* **1992**, *2*, 199; Y. Zeng, Y. T. Pan, N. Asano, R. J. Nash, A. D. Elbein, *Glycobiology* **1997**, *7*, 297, and ref. cit. therein; R. Pal, G. M. Hoke, M. G. Sarngadharan, *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 3384; V. A. Johnson, B. Walker, M. A. Barlow, T. J. Paradis, T. C. Chou, M. S. Hirsch, *Antimicrob. Agents Chemother.* **1989**, *33*, 53; A. Mehta, X. Lu, T. M. Block, B. S. Blumberg, R. A. Dwek, *Proc. Natl. Acad. Sci. U.S.A.* **1997**, *94*, 1822; P. B. Fischer, G. B. Karlsson, R. A. Dwek, F. M. Platt, *J. Virol.* **1996**, *70*, 7153; E. Fenouillet, M. J. Papandreu, I. M. Jonest, *Virology* **1997**, *231*, 89; S. L. White, T. Nagai, S. K. Akiyama, E. J. Reeves, K. Grzegorzewski, K. Olden, *Cancer Commun.* **1991**, *3*, 83; K. Olden, P. Breton, K. Grzegorzewski, Y. Yasuda, B. L. Gause, O. A. Oredipe, S. A. Newton, S. L. White, *Pharmacol. Ther.* **1991**, *50*, 285; P. E. Goss, J. Baptiste, B. Fernandez, M. Baker, J. D. Dennis, *Cancer Res.* **1994**, *54*, 1450.
- [8] K. Kraehenbuehl, S. Picasso, P. Vogel, *Helv. Chim. Acta* **1998**, *81*, 1439.
[9] B. A. Johns, Y. T. Pan, A. D. Elbein, C. R. Johnson, *J. Am. Chem. Soc.* **1997**, *119*, 4856.
[10] C. R. Johnson, M. W. Miller, A. Golebiowski, H. Sundram, M. B. Ksebati, *Tetrahedron Lett.* **1994**, *35*, 8991.
[11] O. R. Martin, L. Li, Y. Feng, *Tetrahedron Lett.* **1996**, *37*, 1991.
[12] M. A. Leeuwenburgh, S. Picasso, H. S. Overkleef, G. A. van der Marel, P. Vogel, J. H. van Boom, *Eur. J. Org. Chem.* **1999**, 1185.
[13] A. Baudat, P. Vogel, *Tetrahedron Lett.* **1996**, *37*, 483; E. Frérot, C. Marquis, P. Vogel, *Tetrahedron Lett.* **1996**, *37*, 2023.
[14] Y. H. Zhu, P. Vogel, *J. Org. Chem.* **1999**, *64*, 666.
[15] C. Marquis, S. Picasso, P. Vogel, *Synthesis* **1999**, 1441.
[16] F. Cardona, S. Valenza, S. Picasso, A. Goti, A. Brandi, *J. Org. Chem.* **1998**, *63*, 7311.
[17] Y.-H. Zhu, P. Vogel, *J. Chem. Soc., Chem. Commun.* **1999**, 1873.
[18] Z. J. Witczak 'Levoglucosenone and Levoglucosans; Chemistry and Applications', ATL Press, Inc., Science Publishers, Mount Prospect, IL, USA, 1994; Z. J. Witczak, *Stud. Nat. Prod. Chem.* **1994**, *14*, 267; Y. Tsuchiya, K. Suami, *J. Appl. Polym. Sci.* **1970**, *14*, 2003; F. Shafizadeh, P. P. S. Chin, *Carbohydr. Res.* **1976**, *46*, 149; F. Shafizadeh, R. H. Fumeaux, T. T. Stevenson, *Carbohydr. Res.* **1979**, *71*, 169; M. Shibagaki, K. Takahashi, H. Kuno, I. Honda, H. Matsushita, *Chem. Lett.* **1990**, 307.
[19] G. W. J. Fleet, J. C. Son, *Tetrahedron* **1988**, *44*, 2647.
[20] K. Takai, M. Tagashira, T. Kuroda, K. Oshima, K. Utimoto, H. Nozaki, *J. Am. Chem. Soc.* **1986**, *108*, 6048; D. P. Stamos, C. Sheng, S. S. Chen, Y. Kishi, *Tetrahedron Lett.* **1997**, *38*, 6355, and ref. cit. therein.
[21] G. Stork, M. Isobe, *J. Am. Chem. Soc.* **1975**, *97*, 6260.
[22] N. T. Anh, *Top. Curr. Chem.* **1980**, *88*, 145; N. T. Anh, T. T. Bui, *Nouv. J. Chim.* **1986**, *10*, 681; see also G. Cainelli, D. Giacomini, P. Galletti, A. Marini, *Angew. Chem., Int. Ed.* **1996**, *35*, 2849; P. Ciapetti, M. Falorni, M. Taddei, *Tetrahedron* **1996**, 7379; F. Ruebsam, S. Seck, A. Giannis, *Tetrahedron* **1997**, *53*, 2823.
[23] T. Imamoto, M. Ono, *Chem. Lett.* **1987**, 501; A. R. Muci, R. Stürmer, D. A. Evans, *J. Org. Chem.* **1993**, *58*, 5307.
[24] D. A. Evans, G. C. Fu, A. H. Hoveyda, *J. Am. Chem. Soc.* **1992**, *114*, 6671.
[25] G. W. Kabalka, S. Yu, N. Li, *Tetrahedron Lett.* **1997**, *38*, 5455.
[26] K. S. Kim, W. A. Szarek, *Synthesis* **1978**, 48.
[27] R. Demange, P. Vogel, in preparation.

Received June 26, 2001